Systematic Gene Search in the Incyte LifeSeq Database

Normal tissue ~50,000 individual ESTs

Tumor tissue ~50,000 individual ESTs

Priority list High

Prostate Breast

Ovary Bladder Uterus Iterative assembling

with

increasing mismatch

Low

~8,000 contigs + ~25,000 individual sequences

Comparison of databases

normal tissuespecific

(expected: 100-500)

nonspecifically expressed genes

tumor tissuespecific (expected: 100-500)

Genes of Interest

Figure 1

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Systematische Gen-Suche in der Incyte LifeSeq Datenbank

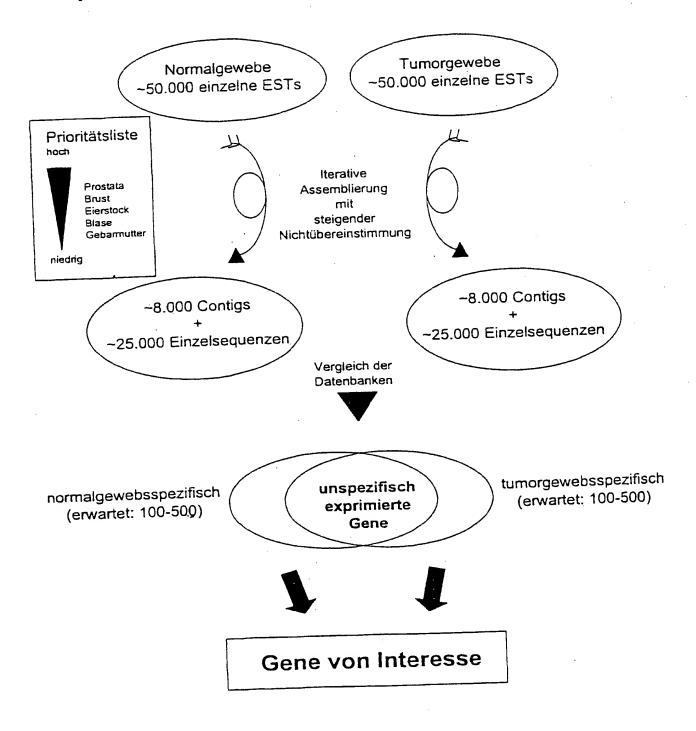


Fig. 1

Principle of EST Assembly ~50,000 ESTs per tissue

Assembly at 0% mismatch with GAP4 (Staden)

Contigs

Individual Sequences

Contigs increasing in number and length

Iterative assembly with increasing mismatch (1%, 2%, 4%)

5000-6000 contigs

~25,000 other individual sequences

~30,000 consensussequences per tissue

Figure 2a

Prinzip der EST-Assemblierung

~50.000 ESTs pro Gewebe



In Anzahl und Länge zunehmenden Contigs

Terative Assemblierung mit steigender Nichtübereinstimmung (1%, 2%, 4%)

5000-6000 Contigs

-25.000 übrige Einzelsequenzen

~30.000 Konsensussequenzen pro Gewebe

Fig. 2a
BERICHTIGTES BLATT (REGEL 91)
ISA/EP

~50,000 ESTs of a tissue (e.g.: uterus tumor)

GAP4 Assembly 1st Round:
 minimum initial match: 20
maximum number of inserted blanks per sequence: 8
 maximum percent mismatch: 0

GAP4 Database 1 Contigs 1 Individual sequences 1 unassembled ESTs

GAP4 Assembly 2nd Round:
 minimum initial match: 20
maximum number of inserted blanks per sequence: 8
 maximum percent mismatch: 1

GAP4 Database 2 Contigs 2 Individual sequences 2 unassembled ESTs

GAP4 Assembly 3rd Round:
 minimum initial match: 20
maximum number of inserted blanks per sequence: 8
 maximum percent mismatch: 2

GAP4 Database 3: Contigs 3 Individual sequences 3 unassembled ESTs

Figure 2b1

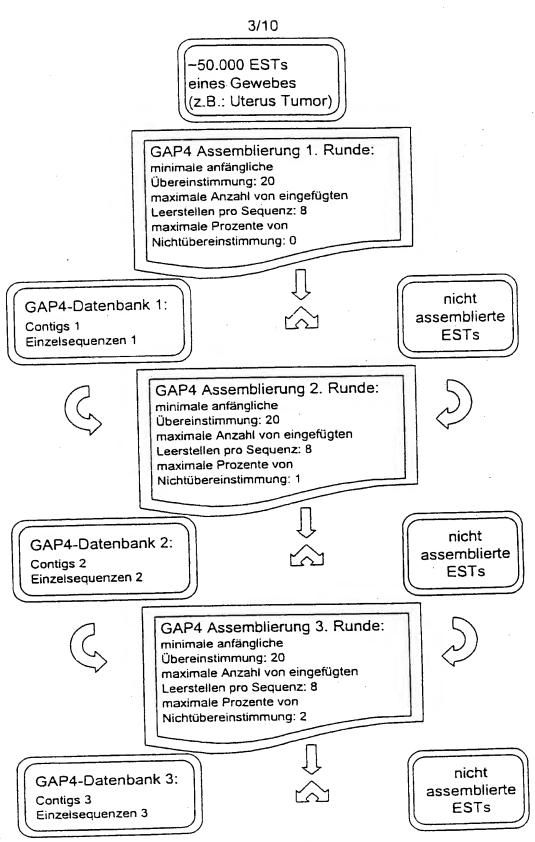


Fig. 2b1

GAP4 Database 3: Contigs 3 Individual Sequences 3 unassembled ESTs

Consensus 3

GAP4 Assembly 4th Round:
minimum initial match: 20
maximum number of inserted blanks
per sequence: 8
maximum percent mismatch: 2

GAP4 Database 4: Contigs 4 Individual Sequences 4 unassembled ESTs

Consensus 4

GAP4 Assembly 5th Round:
minimum initial match: 20
maximum number of inserted blanks
per sequence: 8
maximum percent mismatch: 4

GAP4 Database 5: Contigs 5 Individual Sequences 5 unassembled ESTs 5

Consensus 5

Individual Sequences 5

Figure 2b2

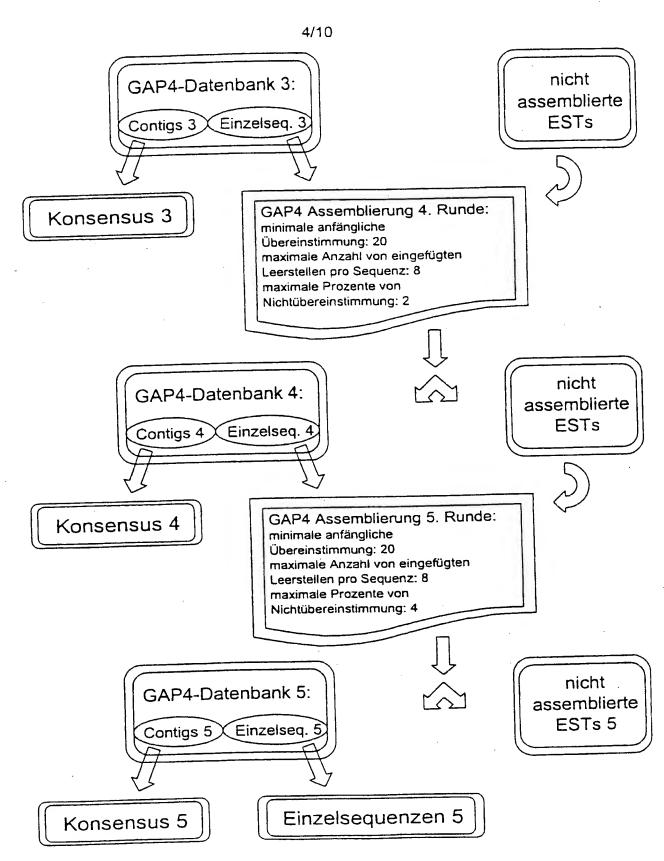


Fig. 2b2

Consensus 3.

Individual Sequences 5

Consensus 4

unassembled ESTs 5

Consensus 5

GAP4 Assembly 6th Round:
minimum initial match: 20
maximum number of inserted blanks per sequence: 8
maximum percent mismatch: 4

Assembled database of a specific tissue (e.g.: uterus tumor)

Figure 2b3

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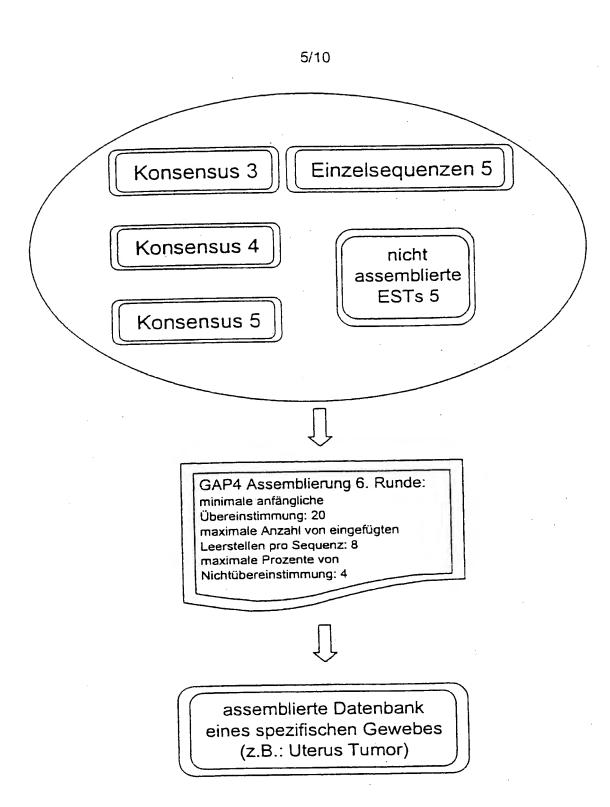


Fig. 2b3

Assembled database of a specific tissue (e.g.: uterus tumor)

Consensus 6

Read-in as individual sequences

Database
of a specific tissue
(e.g.: uterus tumor)

Database of a second specific tissue (e.g.: normal uterus)

GAP4 Assembly minimum initial match: 20 maximum number of inserted blanks per sequence: 8 maximum percent mismatch: 4

Tumor tissuespecific ESTs Non-tissuespecific ESTs Normal tissuespecific ESTs

Fig. 2b4

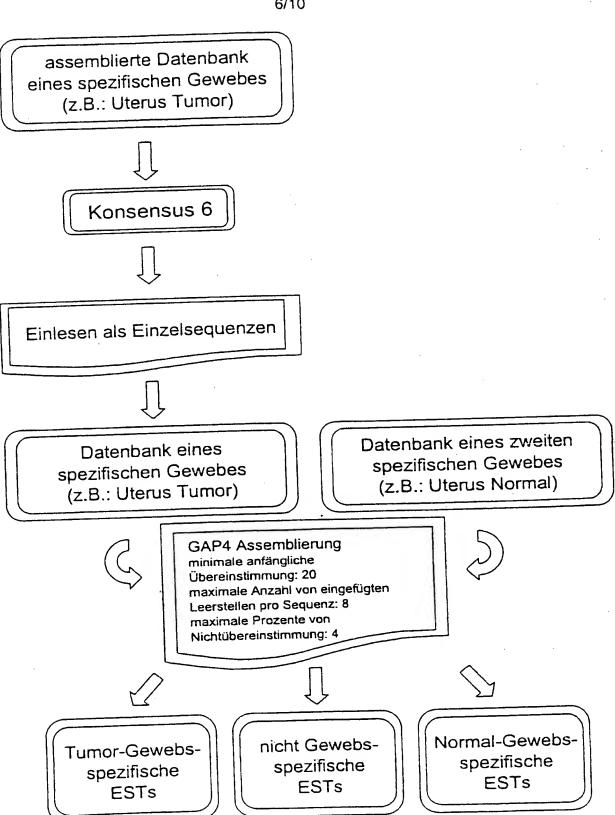


Fig. 2b4

BERICHTIGTES BLATT (REGEL 91) ISA/EP

In silico subtraction of gene expression in various tissues

normal tissue

~30,000 consensus sequences ~30,000 consensus sequences tumor tissue

Assembly at 4% mismatch

Normal tissue Specific genes Cancer tissue Specific genes

Genes expressed in both tissues

Figure 3

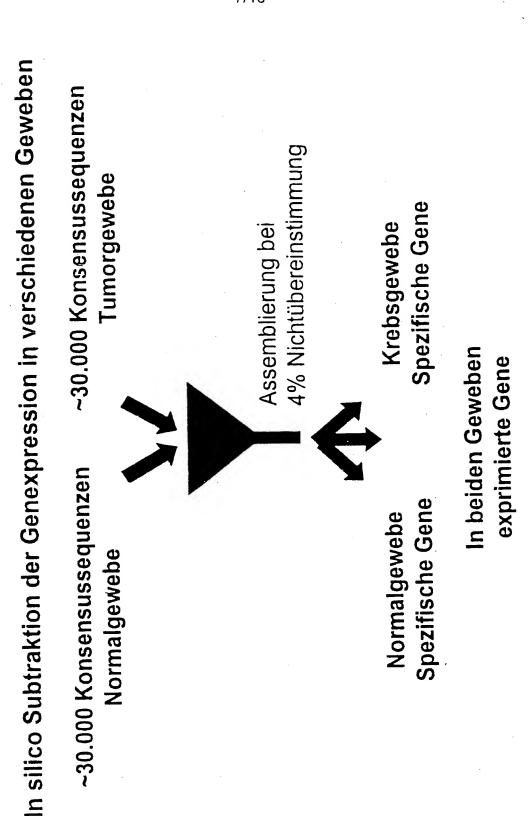


Fig. 3
BERICHTIGTES BLATT (REGEL 91)
154/FP

Genes of interest

Determination of tissue-specific expression via electronic Northern (INCYTE LifeSeq and public EST databases)

Candidate genes for tumor suppressors or tumor activators

Figure 4a

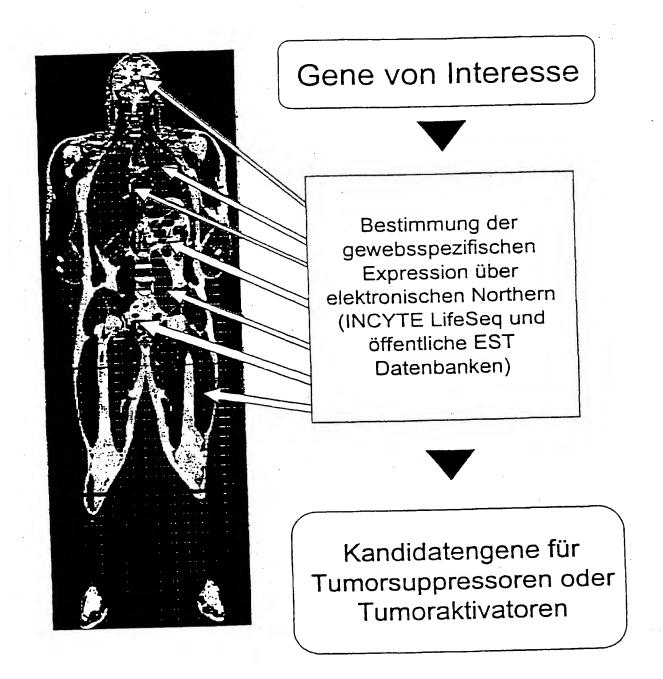


Fig. 4a

Partial cDNA sequence e.g., EST or contig S

...GCCTCAAGTTATC...

WHILE $C_i > C_{i-1}$

Electronic Northern Blot

Fisher's Exact Test $\mbox{ IF }\mbox{ }\mbox$

EXIT

Automatic Lengthening

Consensus sequence C

...ATGTCCTAGCCTCAAGTTATCAGATGCAA...

Figure 4b

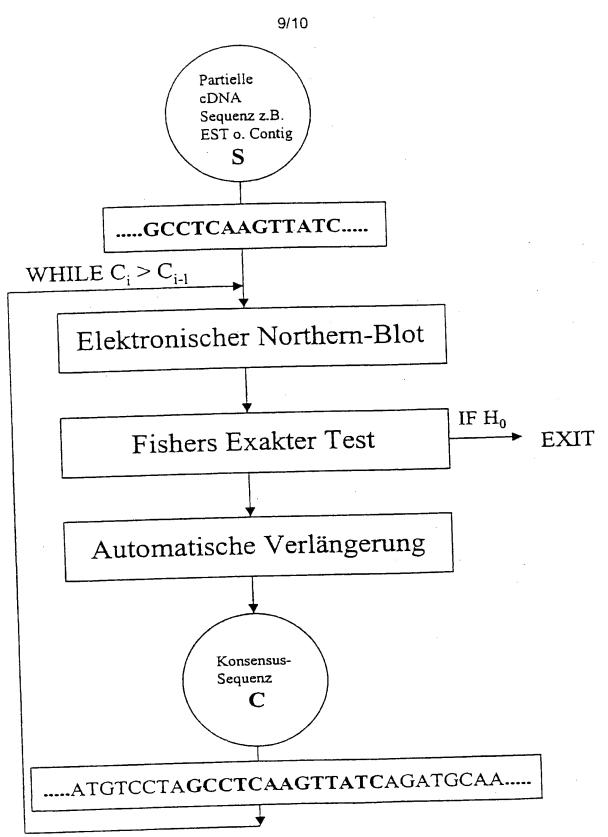


Fig. 4b

Isolation of genomic BAC and PAC clones

Chromosomal clone localization via FISH

Hybridization signal

Sequencing of clones that are located in regions that have chromosomal deletions in prostate and breast cancer leads to identification of candidate genes

Exon Intron

Confirmation of candidate genes by screening of mutations and/or deletions in cancer tissues

Figure 5



Isolieren von genomischen BAC und PAC Klonen



Chromosomale Klon-Lokalisation über FISH



Hybridisierungssignal





Sequenzierung von Klonen, die in Regionen lokalisiert sind, die chromosomale Deletionen in Prostata- und Brustkrebs aufweisen, führt zur Identifizierung von Kandidatengenen

Exon Intron



Bestätigung der Kandidatengene durch Screening von Mutationen und/oder Deletionen in Krebsgeweben

Fig. 5